3-D PSF Fitting for Fluorescence Microscopy: Implementation and Localization Applications

Hagai Kirshner¹, François Aguet², Daniel Sage¹, and Michael Unser¹

Index Terms

Single molecule localization microscopy, 3-D PSF models.

Abstract

Localization microscopy relies on computationally efficient Gaussian approximations of the point spread function (PSF) for the calculation of fluorophore positions. Theoretical predictions show that under specific experimental conditions, localization accuracy is significantly improved when the localization is performed using a more realistic model. Here, we show how this can be achieved by considering 3-D PSF models for the widefield microscope. We introduce a least-squares PSF fitting framework that utilizes the Gibson and Lanni model and propose a computationally efficient way for evaluating its derivative functions. We demonstrate the usefulness of the proposed approach with algorithms for particle localization and defocus estimation, both implemented as plugins for ImageJ.

I. INTRODUCTION

Localization-based fluorescence microscopy relies on sparse activation of individual fluorophores within a sample (Betzig et al., 2006; Hess et al., 2006; Rust et al., 2006). The activated fluorophores are spatially well separated and can be imaged individually. This activate-andimage process is then repeated over many frames, after which the coordinates of each detected

¹Biomedical Imaging Group, École Polytechnique Fédérale de Lausanne (EPFL), Switzerland. ² Department of Cell Biology, Harvard Medical School, Boston MA 02115.

The corresponding author is Hagai Kirshner, phone: +41 (0)21 693.11.36, fax: +41 (0)21 693.68.10, email: hagai.kirshner@epfl.ch, web: http://bigwww.epfl.ch/, postal address: EPFL STI IMT LIB, BM 4142 (Bâtiment BM), Station 17, CH-1015 Lausanne

This work was funded (in part) by the Euro-BioImaging project.

fluorophore are determined computationally and combined to yield the final super-resolved image. The PSF model of the microscope plays a key role in these techniques. Every point-source fluorophore gives rise to a PSF pattern in the image domain, and a localization procedure is applied to the individual patterns. The PSF model that is being used for the localization task determines the accuracy that can be achieved in describing the examined biological structure (Manley et al., 2008; Hedde et al., 2009; Märki et al., 2010; Geissbühler et al., 2011).

Localization accuracy is also determined by the level and type of noise. Poisson noise may appear in the acquired image due to the photon emission characteristics of the fluorophore and due to scattering background noise. Gaussian additive noise, introduced by the imaging sensors, may further reduce the localization accuracy. This matter has been investigated within the context of estimation theory, giving rise to Cramér-Rao lower bounds on the achievable localization accuracy of the Gaussian, the Airy pattern and the Gibson and Lanni models (Ober et al., 2004; Aguet et al., 2005). Many of the currently available localization algorithms utilize the Gaussian model (Bobroff, 1986; Betzig et al., 2006; Hess et al., 2006; Rust et al., 2006; de Moraes Marim et al., 2008; Hedde et al., 2009; Henriques et al., 2010; Wolter et al., 2010).

The Gaussian function provides a reasonable approximation of the main lobe of the Airy pattern while introducing relatively low computational complexity. Such approximation, however, discards the side-lobes of the PSF, which are particularly important in 3-D PSF modeling (Zhang et al., 2007). The trade-off between choosing realistic- and simplified PSF models is execution time, and we propose here to apply a two-stage approach: fast algorithms that rely on simplified PSF models can be used to obtain preliminary results as well as immediate feedback about the quality of the experiment while more realistic 3-D PSF models can be used for a more accurate analysis, performed at a later stage.

In this work we introduce a least-squares PSF fitting framework that utilizes realistic 3-D PSF models. In particular, the Gibson and Lanni model was shown to be very useful for restoration problems in microscopy (Markham and Conchello, 2001; Preza and Conchello, 2004), and we demonstrate its usefulness for particle localization and for defocus estimation, too. The least-squares localization approach is likely to yield less accurate results than the maximum-likelihood approach in the presence of non-Gaussian noise sources (Aguet, 2009), and a quantitative comparison of these two criteria was carried out in Abraham et al. (2009) for the Gaussian and for the Airy disk patterns. It was shown there that in terms of performance, the least-squares fitting

method follows the maximum-likelihood method quite closely, introducing standard deviations that are larger by no more than 2[nm] for the estimated lateral position of a particle. An exception to that is the case of relatively strong mismatch between the width values of the simulated and the fitted PSFs. This can, however, be taken into account by estimating this parameter from the data itself, or by optimizing for it, too.

These findings make the least-squares criterion an attractive and nearly optimal method for PSF fitting tasks. It is a simple yet powerful tool that depends on the fitted model only. Its additional advantage is that it lends itself to a fast minimization using the Levenberg-Marquardt algorithm. The maximum-likelihood criterion, on the other hand, requires additional knowledge on the noise sources and relies on optimization procedures that are in many cases more involved in terms of the cost function and in terms of the numerical implementation of the minimization procedure (Aguet et al., 2005; Abraham et al., 2009).

The paper is organized as follows: in Section II we describe the Gibson and Lanni model and compute its partial derivative functions while taking into account the stage displacement, the particle axial position and the defocus measure of the detector plane. In section III we introduce an efficient way of evaluating these functions. In Section IV we utilize the 3-D PSF model for localizing particles in a z-stack. We fit the data with the 3-D position coordinates and with an amplitude value that accounts for the random nature of the photon emission rate. Our algorithm uses adaptive threshold values for local maxima identification, and an adaptive window size for the least-squares fit. Motivated by multi-plane imaging (Prabhat et al., 2004; Ram et al., 2008), we also introduce an algorithm for estimating the defocus distance of the detector plane. All of our algorithms were implemented as ImageJ plugins¹; they are briefly described in the Appendix.

II. GIBSON AND LANNI MODEL

The Gibson and Lanni model generalizes the Born and Wolf model by accommodating for a refractive index mismatch between the three imaging layers (Appendix A). It assumes an optical path that includes a biological sample, a coverslip layer and an immersion layer (Figure 1). It relies on the Li and Wolf approximation of the Kirchhoff diffraction integral

$$h\left(\theta\right) = \left(\frac{ka^{2}A_{0}}{z_{d}^{2}}\right)^{2} \left| \int_{0}^{1} J_{0}\left(\frac{kar(\theta)\rho}{z_{d}}\right) e^{iW(\rho;\theta)}\rho \,d\rho \right|^{2},\tag{1}$$

¹The software is available at http://bigwww.epfl.ch/algorithms/psfgenerator



Fig. 1. The Gibson and Lanni model assumes three layers for the optical path.

where θ is a set of parameters given in Table I, J_0 is the Bessel function of the first kind of order zero and $k = 2\pi/\lambda$ is the wave number of the emitted light. *a* is the radius of the circular aperture at the back focal plane and it can be approximated by

$$a \cong \mathrm{NA}z_d^*/M. \tag{2}$$

 $W(\rho; \theta)$ describes the optical path difference

$$W(\rho;\theta) = kn_s z_p \sqrt{1 - \left(\frac{\mathrm{NA}\rho}{n_s}\right)^2} + kn_i \Delta t_i \sqrt{1 - \left(\frac{\mathrm{NA}\rho}{n_i}\right)^2} + \frac{ka^2(z_d^* - z_d)}{2z_d^* z_d} \rho^2, \tag{3}$$

and $r(\theta)$ is the lateral distance between the particle position and the detector in the image domain

$$r(\theta) = \sqrt{(x_p - x_d)^2 + (y_p - y_d)^2}.$$
(4)

The original expression of $W(\rho; \theta)$ in Gibson and Lanni (1992) distinguishes between nominal and actual refractive indices values of the immersion layer. Here, we assume that they are both equal, and nominal conditions are also applied to the coverslip thickness value. The Gibson and Lanni model assumes a homogenous sample layer and this is one of its limitations. Variations in refractive index values can be measured by differential interference contrast techniques (Kam et al., 2001) or be modeled as a stochastic process (Schmitt and Kumar, 1996). This means that $W(\rho; \theta)$ is no longer a deterministic function, so one can interpret n_s as an effective refractive index value. The optical path difference (3) can be alternatively expressed by means of the defocus measure in the object space (Aguet et al., 2005). The advantage of expressing $W(\rho; \theta)$ by means of stage displacement values, as done here, is the straight-forward calculations for z-stack acquisitions. We also included a defocus measure in the image space, allowing one to compute the PSF pattern of the multi-plane design (Prabhat et al., 2004).

One of the advantages of the The Gibson and Lanni model is its ability to predict nonsymmetric PSF patterns. Such patterns occur due to refractive index mismatch between the sample layer and the immersion layer (Figure 2). Another advantage is the distinction between stage displacements, detector location and particle depth. Each parameter has a different effect in terms of defocusing. The defocus measure can be approximated by the value of the Taylor coefficient of ρ^2 in $W(\rho; \theta)$, giving rise to the following relation

$$\frac{z_p}{n_s} + \frac{\Delta t_i}{n_i} = \frac{a^2 (z_d^* - z_d)}{NA^2 z_d^* z_d}.$$
(5)

This criterion implies that focusing can be achieved by either moving the stage, the detector, or the particle itself.

III. NUMERICAL EVALUATION AND FITTING

We consider the task of fitting the Gibson and Lanni PSF model to light patterns that originate from single point sources. We minimize the least-squares criterion by using the Levenberg-Marquardt method. To this aim we express (1) as follows

$$h(\theta) = C(\theta) \left[\left(I_1(\theta) \right)^2 + \left(I_2(\theta) \right)^2 \right], \tag{6}$$

where

$$C(\theta) = \left(\frac{ka^2 A_0}{z_d^2}\right)^2 \tag{7}$$

$$I_1(\theta) = \int_0^1 J_0\left(\frac{kar(\theta)\rho}{z_d}\right) \cos\left(W(\rho;\theta)\right) \rho \,d\rho,\tag{8}$$

$$I_2(\theta) = \int_0^1 J_0\left(\frac{kar(\theta)\rho}{z_d}\right) \sin\left(W(\rho;\theta)\right) \rho \,d\rho.$$
(9)

This, in turn, allows one to write

$$\frac{\partial h(\theta)}{\partial \upsilon} = \frac{\partial C(\theta)}{\partial \upsilon} \left[\left(I_1(\theta) \right)^2 + \left(I_2(\theta) \right)^2 \right] + C(\theta) \left[2I_1 \frac{\partial I_1(\theta)}{\partial \upsilon} + 2I_2 \frac{\partial I_2(\theta)}{\partial \upsilon} \right], \tag{10}$$



Fig. 2. A cross-section the Gibson and Lanni PSF pattern. The PSF parameters θ are: NA = 1.4, $n_i = 1.5$, $n_s = 1.33$, $\lambda = 520$ [nm], $x_p = y_p = 0$, $z_p = 1000$ [nm], $z_d = z_d^*$, $x_d = 0$. The horizontal axis is Δt_i and the vertical one is y_d . The upper left corner is ($\Delta t_i, y_d$) = (-12775, -12275) in [nm] and both the pixel size and the z-step value are 50[nm]. Pixel values are in the range [0, 1] and the saturation level was set to 0.001 (left) and to 0.1 (right) as to demonstrate the non-symmetric nature of the PSF due to the refractive index mismatch. The focal plane corresponds to $\Delta t_i = -1350$ which is approximately $z_p n_i/n_s$.

where v is one of the parameters of θ . The work of Ram et al. (2005) introduces generalized expressions for the Fisher information matrix with respect to the 3D particle position. Starting from the Kirchhoff diffraction integral (1), the authors computed the partial derivative of the PSF as a function of $W(\rho; \theta)$. We apply these results to the Gibson and Lanni model and provide in Table II explicit expressions for $\partial I_1(\theta)/\partial v$, $\partial I_2(\theta)/\partial v$.

To evaluate such integrals accurately, one needs to adapt the integration step to the oscillatory nature of the integrands. For example, as the particle is located deeper into the sample, the integrands in Table II oscillate more rapidly, requiring more sampling points for the numerical approximation. A possible approach for the integral calculation was suggested in Aguet et al. (2005). There, the number of sampling points was determined by the highest possible oscillation rate of either the optical path difference or the Bessel function. Such a Nyquist-based approach assumes that the integrands are essentially band-limited, although they are better modeled as

chirp functions. For this reason, the band-width values of the integrands are relatively large and so is the required number of sampling points. We therefore take a different point of view that relies on approximation theory.

We approximate the integrals in a progressive manner. Let the sampling interval at the *n*-th iteration be $(1/2)^n$, and let $\hat{h}_n(\theta)$ be the approximated value for $h(\theta)$. The integrands that appear in I_1, I_2 are smooth functions and we approximate them by a piecewise quadratic function using the Simpson method. This means that the approximation error $\epsilon_n(\theta) = h(\theta) - \hat{h}_n(\theta)$ has a certain rate of decay. In particular, there exists N > 0 for which for all n > N

$$|\epsilon_n(\theta)| \le D(\theta) \left(2^{-n}\right)^L.$$
(11)

 $D(\theta)$ does not depend on *n*, nor does the decay rate *L* which reflects the approximation order of the Simpson method. We define

$$\eta_n(\theta) = \hat{h}_{n-1}(\theta) - \hat{h}_n(\theta), \tag{12}$$

and observe that

$$|\eta_n(\theta)| = |\epsilon_{n-1}(\theta) - \epsilon_n(\theta)|$$
(13)

$$\leq \frac{3}{2}D(\theta)\left(2^{-n}\right)^{L}.$$
(14)

This means that $\eta_n(\theta)$ has a decay rate that is not larger than the decay rate of $\epsilon_n(\theta)$. As L and $D(\theta)$ are not known in practice, we extract them from $\eta_n(\theta)$ as demonstrated in Figure 3. In particular, we can find $\tilde{D}(\theta)$ and \tilde{L} such that

$$|\eta_n| \le \tilde{D}(\theta) \left(2^{-n}\right)^{\tilde{L}}.$$
(15)

Now,

$$|\epsilon_n(\theta)| = \left| h(\theta) - \hat{h}_n(\theta) \right|$$
(16)

$$= \left| \sum_{k=n+1}^{\infty} \eta_k(\theta) \right| \tag{17}$$

$$\leq \sum_{k=n+1}^{\infty} \tilde{D}(\theta) \left(2^{-\tilde{L}}\right)^k \tag{18}$$

$$\leq \tilde{D}(\theta) \left(2^{-n}\right)^{\tilde{L}} \frac{2^{-L}}{1 - 2^{-L}}.$$
(19)

We approximate $\tilde{D}(\theta) (2^{-n})^{\tilde{L}}$ by $|\eta_n(\theta)|$ and have $|\epsilon_n(\theta)| \cong |\eta_n(\theta)| \frac{2^{-L}}{1-2^{-L}}$.

In order to express the error in terms of percentage, we use the relative error measure $\frac{|\eta_n(\theta)|}{\hat{h}_n(\theta)}$ as a stopping criterion. The number of iterations is determined by a threshold on the relative error, say 1%. To ensure that the approximation error is governed by the decay rate (11), we require it to meet this threshold for at least three consecutive iterations. The numerical evaluation of the Bessel functions impose no limitation on $\epsilon_n(\theta)$; the Bessel functions J_0 is evaluated up to an accuracy of $5 \cdot 10^{-8}$ and the J_1 function up to $5 \cdot 10^{-8}$ times its argument (Abramowitz and Stegun, 1972).

The advantage of such progressive evaluation resides in the fact that less computational time is spent on calculating integrals of low $W(\rho; \theta)$ values while controlling the accuracy level of their numerical approximation (Figure 4). Extensions to other 3-D PSF designs are possible, too. In particular, to the biplane setup (Prabhat et al., 2004; Juette et al., 2008; Kirshner et al., 2012), to the astigmatic PSF (Huang et al., 2008), and to the double helix pattern (Pavani and Piestun, 2008). The only modification is in the expression of the phase term $W(\rho; \theta)$. The Kirchhoff diffraction integral remains the same so our numerical implementation can still be used for these cases.

IV. APPLICATIONS

We consider two applications: 3-D particle localization, and misalignment estimation. We introduce a least-squares fitting algorithm and analyze its performance in the presence of Poisson and Gaussian noise sources. The description of the software that was developed for this work is described in Appendix B.

A. 3-D particle localization

One possible way of determining the 3-D location of a particle is by acquiring several PSF images that correspond to different sets of PSF parameters θ . A z-stack, for example, provides PSF images with different thickness values of the immersion layer. Another option is to have several detector planes located at different positions along the optical path in the image domain. The former corresponds to different values of Δt_i while the latter to different values of z_d . Assuming a z-stack of fixed point sources, we introduce the following algorithm (Figure 5).

Normalization: The z-stack is normalized to have a unit maximum value.



Fig. 3. Converging properties of the PSF numerical approximation. Shown here on the left column are PSF values for two sets of parameters θ_1, θ_2 . The following parameters are common to both of them: NA = 1.4, $n_i = 1.5, n_s = 1.33, \lambda =$ $520[nm], M = 100, x_p = y_p = 0, z_d = z_d^*, y_d = 0$. The point source depth value z_p is 200[nm] for θ_1 and 800[nm] for θ_2 . The detector lateral position x_d is 0 and 1500 [nm] respectively. The right column depicts relative difference values between every two consecutive iterations. All figures indicate logarithmic values, and the decay rate of the relative error is $\tilde{L} \cong 1/\ln 2$.

Local maxima identification: We use different threshold values for the different slices. This is because the amplitude of the focused pattern fades away as the particle moves deeper into the sample. The Δt_i parameter of a given slice provides us with an initial estimation of the particle depth by means of (5). We then calculate a PSF value for $r(\theta) = 0$, denoted $h(\theta_{slice})$, and a PSF value for the global maximum, denoted $h(\theta_{max})$. The ratio between these two PSF values approximates the expected ratio between a local maximum at a particular slice and the global maximum. Deviations from this ratio may occur due to sub-pixel axial and lateral positions, random nature of the emission rate, and noise. For these reasons we choose the threshold to be

$$\tau_{slice} = \frac{1}{2} \cdot \frac{h(\theta_{slice})}{h(\theta_{max})}.$$
(20)



Fig. 4. A cross-section the Gibson and Lanni PSF pattern. Pixel values denote the number of sampling points that were required for calculating the PSF value. The minimum value is 8 (dark) and the maximum value is 65535 (bright). Similar to Figure 2, the PSF parameters θ are: NA = 1.4, $n_i = 1.5$, $n_s = 1.33$, $\lambda = 500$ [nm], $x_p = y_p = 0$, $z_p = 1000$ [nm], $z_d = z_d^*$, $x_d = 0$. The horizontal axis is Δt_i and the vertical one is y_d . The upper left corner is ($\Delta t_i, y_d$) = (-12775, -12275) in [nm] and both the pixel size and the z-step value are 50[nm]. The focal plane corresponds to $\Delta t_i = -1350$ which is approximately $z_p n_i/n_s$.



Fig. 5. Main stages of the localization algorithm.

For large stage displacement values, τ_{slice} may be lower than the mean noise level, and for that reason the local maxima should be higher than the noise level, too. We estimate the mean mand variance σ of the noise from the z-stack itself and set this threshold value to be $m + 3\sigma$. In addition to the voxel value, the local maxima should be higher than its 26 3-D neighbors.

Least-squares fitting: The 3-D neighborhood of each local maxima is fitted with the Gibson and Lanni model, where the size of the fitting window is $2r \times 2r \times 2d$. We set $r = \frac{7}{k} \frac{M}{\text{NA}} \frac{1}{\text{pixel size}}$ to be the distance to the second minimum of the Airy pattern. This distance captures 90% of the energy of the Airy pattern and we use the same criterion for determining the axial parameter d by numerical means. For the PSF parameters of Figure 2, the values of r and d are 8[pixels] and 9[pixels], respectively. Initial values for (x_p, y_p) are the coordinates of the local maximum and the initial value for z_p is given by (5). The initial value for A_0 is the ratio between the pixel value of the local maximum and the value of the PSF model for $(x_d, y_d) = (x_p, y_p)$. Derivative values of the PSF model are given in Table II and (10).

Goodness of fit: We use several criteria for accepting the fitted parameters: the lateral position should be close to the local maximum, the axial position should be close to the initial guess and the SNR (Signal to Noise Ratio) value of the fit should be above a certain threshold.

Simulated z-stack data is shown in Figure 6 and least-squares localization performance is demonstrated in Figures 7–9. The fitting window was $6 \times 6 \times 6$ pixels. Our performance analysis included several particle depth values, several lateral positions, and several noise levels. Each setup was repeated 20 times. Following Ober et al. (2004), the noise sources are Poisson-distributed scatter noise and Gaussian-distributed readout noise. The emission rate of every point source is a Poisson random number. Our results suggest that the lateral localization accuracy is less sensitive to the particle depth value, compared with the axial localization accuracy. The relatively low localization accuracy for the depth value of 200[nm] follows from the smaller number of data points that can be used in the fitting process. This is due to the fact that the z-stack consists of positive stage displacement values only, demonstrating the importance of the fitting window size for the axial localization. We also observe that the sub-pixel lateral positions have minor effect on the lateral and axial localization accuracies.

36400.00x36400.00 nm (364x364); 32-bit; 518K	Z Stack (300%)	
$\Delta t_i = 0$	$\Delta t_i = 400 [nm]$	$\Delta t_i = 800[nm]$
$\Delta t_i = 1200[nm]$	$\Delta t_t = 1600[nm]$	$\Delta t_i = 2000[nm]$
$\Delta t_i = 2400[nm]$	$\Delta t_i = 2800[nm]$	$\Delta t_i = 3200[nm]$ 5000 nm

Fig. 6. Simulated z-stack. Ten particles were randomly located in a 3-D volume and the their PSF image were calculated based on the Gibson and Lanni model for different stage displacement values. The PSF parameters θ are: NA = 1.4, $n_i =$ $1.5, n_s = 1.33, \lambda = 520$ [nm]. The noise parameters are: mean emission rate of $2 \cdot 10^6$ [photons/sec], optical efficiency of 0.033, acquisition time of 0.1[sec], readout rms noise of 6[electrons/pixel], and mean scattering rate of 660[photons/sec]. The depth values of the particle are in the range of [0, 2000][nm].



Fig. 7. Lateral positions within a pixel. Position A is located at the center of the pixel and positions C, E and F are at the boundaries. These sub-pixel positions were used for evaluating the performance of the proposed PSF fitting algorithms.

B. Misalignment estimation

Motivated by the calibration stage of multiplane microscopic design (Prabhat et al., 2004; Juette et al., 2008), we suggest here a fitting algorithm that allows for the determination of the axial position, z_d , of the defocused detector plane. The input data is a z-stack of fixed point-



Fig. 8. Lateral (left) and axial (right) localization accuracy for various particle depth values and for several lateral positions. Every localization accuracy value was computed by averaging over 20 realizations. The acquisition parameters θ are: NA = $1.4, n_i = 1.5, n_s = 1.33, \lambda = 520$ [nm]. The pixel size is 150[nm] and the z-step size is 100[nm]. The noise parameters are: mean emission rate of $2 \cdot 10^6$ [photons/sec], optical efficiency of 0.033, acquisition time of 0.1[sec], readout rms noise of 6[electrons/pixel], and mean scattering rate of 660[photons/sec]. The sub-pixel lateral positions A, B, C, D, E and F are described in Figure 7. The axial localization error (right) is taken in the absolute value sense, as to comply with the positive values of the lateral localization error (left). The highest standard deviation value for the lateral position was 0.2[nm], and for the axial position – 1.4[nm].

sources that lie on the coverslip $z_p = 0$. Stage displacement values are both positive and negative with respect to the working distance. The output of the algorithm are the estimated values of x_p, y_p, z_d and A_0 . The algorithm follows the stages of Figure 5 with the modification that the initial parameter for z_d is determined by sweeping over several possible values and choosing the one that maximizes the PSF value at the particular slice of the z-stack. The threshold value is set to $\tau = 0.5$ accounting for the Poisson distribution of the emission rate and for the noise. If all of the particles lie at the same depth position, then local maxima should appear in the same frame. Due to noise, the local maxima may appear in neighboring slices and this does not require re-calculating τ , as was done in the previous algorithm. Fitting results are depicted in Figures 10-11. They suggest that the least-squares criterion provides an unbiased estimation with a relatively low variance value for the axial position of the detector plane.



Fig. 9. Lateral (left) and axial (right) localization accuracy for various particle depth values and for several lateral positions. Every localization accuracy value was computed by averaging over 20 realizations. The acquisition parameters θ are: NA = $1.4, n_i = 1.5, n_s = 1.33, \lambda = 520$ [nm]. The pixel size is 150[nm] and the z-step size is 100[nm]. The noise parameters differ from the parameters of Figure 8: mean emission rate of $2 \cdot 10^6$ [photons/sec], optical efficiency of 0.033, acquisition time of 0.1[sec], readout rms noise of 36[electrons/pixel], and mean scattering rate of 6600[photons/sec]. The highest standard deviation value for the lateral position was 1.1[nm], and for the axial position - 8.1[nm].



Fig. 10. Lateral (left) and defocus (right) localization accuracy for various defocus values of the detector plane and for several particle lateral positions. Every localization accuracy value was computed by averaging over 20 realizations. The acquisition parameters θ are: NA = 1.4, $n_i = 1.5$, $n_s = 1.33$, $\lambda = 520$ [nm]. The pixel size is 150[nm] and the z-step size is 100[nm]. The nominal value for z_d is $z_d^* = 0.2$ [m]. The noise parameters are: mean emission rate of $2 \cdot 10^6$ [photons/sec], optical efficiency of 0.033, acquisition time of 0.1[sec], readout rms noise of 6[electrons/pixel], and mean scattering rate of 660[photons/sec]. The sub-pixel lateral positions A, B, C, D, E and F are described in Figure 7. The highest standard deviation value for the lateral position was 0.28[nm], and for the axial position $- 0.17 \cdot 10^{-4}$ [m].



Fig. 11. Lateral (left) and defocus (right) localization accuracy for various defocus values of the detector plane and for several particle lateral positions. Every localization accuracy value was computed by averaging over 20 realizations. The acquisition parameters θ are: NA = 1.4, $n_i = 1.5$, $n_s = 1.33$, $\lambda = 520$ [nm]. The pixel size is 150[nm] and the z-step size is 100[nm]. The nominal value for z_d is $z_d^* = 0.2$ [m]. The noise parameters differ from the parameters of Figure 10: mean emission rate of $2 \cdot 10^6$ [photons/sec], optical efficiency of 0.033, acquisition time of 0.1[sec], readout rms noise of 36[electrons/pixel], and mean scattering rate of 6600[photons/sec]. The highest standard deviation value for the lateral position was 2[nm], and for the detector plane position $-1.35 \cdot 10^{-4}$ [m].

V. CONCLUSIONS

In this work, we provided a general approach to 3-D PSF fitting applications, while introducing an efficient and accurate evaluation of the PSF function. We focused on the Gibson and Lani model and showed its usefulness in particle localization and in defocus estimation. Our fitting algorithm relies on the least-squares error measure, and we provided analytical expressions for the required partial derivative functions. We then introduced an efficient and accurate way for evaluating the Kirchhoff diffraction integral. Our fitting algorithm includes local maxima identification with adaptive threshold values and window size for the least-squares fit. The algorithm utilizes the Levenberg-Marquardt minimization method and the initial values we use for it rely on the PSF model. Simulation results with z-stack data indicate that the lateral localization accuracy is less sensitive to the particle depth value, compared with the axial localization accuracy. It was also shown that axial localization accuracy can be improved substantially by taking more data points in the axial direction. We also observed that sub-pixel lateral positions have minor effect on the lateral and axial localization accuracies when using the least-squares criterion. Our results also suggest that this criterion provides an unbiased estimation with a relatively low variance value for the axial position of the detector plane.

APPENDIX

A. The relation between the Gibson – Lanni and the Born–Wolf models

The Born and Wolf model is

$$h\left(\theta\right) = \left(\frac{ka^{2}A_{0}}{z_{d}^{2}}\right)^{2} \left| \int_{0}^{1} J_{0}\left(\frac{kar(\theta)\rho}{z_{d}}\right) e^{\frac{-ika^{2}\rho^{2}}{2z_{d}^{2}}\left(z_{d}^{*}-z_{d}\right)}\rho \,d\rho \right|^{2}.$$
(21)

We set the aperture, a, to be the back focal plane of the microscope and rely on the sine condition $n \sin \theta = M \sin \phi$ to have the following relation (Figure 12)

$$NA = n \sin \theta_{max} = M \sin \phi_{max} = \frac{Ma}{\sqrt{a^2 + (z_d - f)^2}},$$
 (22)

The parameter z_d denotes the distance of the focused image from the back principle plane, and z_d^* denotes the distance of the detector from the same plane. In practice, $z_d \gg f$ and $M \gg NA$ which leads to

$$\frac{a}{z_d} \cong \frac{\mathrm{NA}}{M}.$$
(23)

We express the defocusing measure $\Delta z = z_d^* - z_d$ in the image domain in terms of defocusing $\Delta z' = z_d' - z_d^{*'}$ in the object domain by relying on the analysis of Gibson (Gibson, 1990, Chapter 3). $z_d^{*'}$ is the distance from the front focal plane for which a particle would be focused on the detector plane z_d^* , i.e. just beneath the coverslip. z_d' is the distance for which a particle will be focused at z_d in the image domain. The two pairs of similar triangles in Figure 13 yield

$$\frac{z_d'}{f'} = \frac{f}{z_d} \cong \frac{1}{M},\tag{24}$$

where $f' = n \cdot f$. Assuming small defocusing values, $\Delta z' \ll z'_d$,

$$\Delta z = z_d^* - z_d$$

$$= f f' \left(\frac{1}{z_d^{*'}} - \frac{1}{z_d'} \right)$$

$$= \frac{(f')^2}{n} \left(\frac{z_d' - z_d^{*'}}{z_d^{*'} z_d'} \right)$$

$$\cong \frac{M^2}{n} \Delta z'.$$
(25)



Fig. 12. Geometric description of the sine law in a microscope $n \sin \theta = M \sin \phi$. f is the focal length of the microscope in the image domain and f' is the focal length in the object domain. a is the radius of the circular aperture of the microscope at the back focal plane. z_d is the distance of the detector plane from the back principle plane and z'_d is the distance of a point-source from the front focal plane for which its focused image will located at the detector plane. It is assumed here that the immersion layer, the coverslip and the sample layer share the same refractive index value n.

Substituting (23) and (25) in (21) results in

$$h\left(\theta\right) = \left(\frac{ka^{2}A_{0}}{z_{d}^{2}}\right)^{2} \left| \int_{0}^{1} J_{0}\left(\frac{kNAr(\theta)\rho}{M}\right) e^{\frac{-ikNA^{2}\rho^{2}\Delta z'}{2n}} \rho \, d\rho \right|^{2}, \tag{26}$$

which is the Born and Wolf model for the wide field microscope.

The Gibson and Lanni model generalizes the model of Born and Wolf in the following manner. Assume that all the layers have the same refractive indices $n_s = n_i = n$. It then follows that $OPD(\rho) = n\Delta z' \left[1 - (NA\rho/n)^2\right]^{\frac{1}{2}}$ where the defocus measure in the object domain is due to the particle's depth and stage displacement $\Delta z' = z_p + \Delta t_i$. The Taylor series for the OPD is

$$OPD(\rho) = n\Delta z' - \frac{NA^2 \Delta z \ \rho^2}{2n} - \frac{NA^2 \Delta z \ \rho^4}{8n^3} + O(\rho)^6,$$
(27)

and the coefficient of ρ^2 which amounts to defocusing coincides with the phase term of (26).

B. Software description

The Gibson and Lanni PSF model was implemented in Java, and it can be used with a variety of software packages. As an example application, we developed the ImageJ plugin PSFGenerator, which evaluates and visualizes the 3D pattern of the Gibson and Lanni model². It is fast and easy

²The software is available at http://bigwww.epfl.ch/algorithms/psfgenerator



Fig. 13. The Gaussian image of a defocused object in a widefield microscope. $z_d^{*'}$ denotes the nominal location of an object for which its focused image is located at the detector plane z_d^* . These values are measured relative to the front and back principle planes, respectively. z_d' is the axial position of a particle for which its focused image is located at z_d . f and f' are the focal lengths at the image domain and at the object domain , respectively. They are given by f' = nf where n is the refractive index of the object domain. The lateral magnification is M; drawing not shown to scale.

to use, requiring only few input parameters that are readily available for microscopy practitioners. The output of the plugin is a z-stack of any chosen size, which can also be visualized with orthogonal views. The plugin also provides a table that calculates the maximum value, the proportional energy, and the effective radius of each slice (Figure 14). The open architecture of the plugin allows for easy incorporation of additional PSF models, and the current version includes the Born & wolf and the Richards & Wolf models.

We also used our Gibson and Lanni Java implementation for simulating datasets of photoactivated fluorophores³. To this aim, a user-dependent biological structure is described by a 3-D density map of fluorophores (Figure 15). Every fluorophore is assigned a random position (based on the density map), a random excitation time instant, and a random photon emission rate value. We then use the Gibson and Lanni model to determine the image of each fluorophore and add the image to the time frame it belongs to. Background scatter noise and readout noise are added, too. The output of our software is a sequence of frames, composed of images of photo-activated fluorophores. We also provide the $(x_p, y_p, z_p, frame number)$ indices of every fluorophore, which

³The software is available at http://bigwww.epfl.ch/palm/



Fig. 14. Screen shot of our ImageJ plugin that generates PSF models. Shown are the interface (right top), the z-stack and its orthogonal views (left top), as well as the statistical analysis of the PSF (right bottom). The acquisition parameters are given in the interface itself.

can be used as a validation tool for other localization algorithms.



Fig. 15. An example of a simulated photo-activated fluorophores sequence. The 3-D structure (a) is described by a density map (b, shown is a maximum z-projection). Point sources are then generated based on the density map (c). Every point-source is then excited at a random time frame, giving rise to a set of frame sequence (d). We use the Gibson and Lanni model to determine the image of each fluorophore and we also account for background scatter and readout noise sources.

TABLE I Parameters of the Gibson and Lanni model

Name	Description
NA	Numerical aperture of the microscope
n_s	Refractive index of the specimen layer
n_i	Refractive index of the immersion layer
λ	Emission wavelength in vacuum
Δt_i	Stage displacement relative to the nominal working distance of the objective lens, i.e. axial step size
A_0	average magnitude of the spherical wave that impinges on the back focal plane of the microscope
x_p, y_p	Lateral position of the point source relative to the optical axis
z_p	Axial location of the point-source fluorophore in the specimen layer relative to the coverslip
x_d, y_d	Lateral position of a pixel in the image domain relative to the optical axis
z_d, z_d^*	Axial distance of the detector plane from the back principle plane. The nominal value z_d^* can be approximated by
	the tube length value of the microscope

DERIVATIVE EXPRESSIONS FOR THE GIBSON AND LANNI PSF MODEL

_

Parameter	Derivative expressions
	$\frac{\partial I_1(\theta)}{\partial x_p} = \frac{ka}{z_d} \frac{x_d - x_p}{r(\theta)} \int_0^1 J_1\left(\frac{kar(\theta)\rho}{z_d}\right) \cos\left(W(\rho;\theta)\right) \rho^2 d\rho$
x_p	$\frac{\partial I_2(\theta)}{\partial x_p} = \frac{ka}{z_d} \frac{x_d - x_p}{r(\theta)} \int_0^1 J_1\left(\frac{kar(\theta)\rho}{z_d}\right) \sin\left(W(\rho;\theta)\right) \rho^2 d\rho$
	$\frac{\partial C(\theta)}{\partial x_p} = 0$
	$\frac{\partial I_1(\theta)}{\partial y_p} = \frac{ka}{z_d} \frac{y_d - y_p}{r(\theta)} \int_0^1 J_1\left(\frac{kar(\theta)\rho}{z_d}\right) \cos\left(W(\rho;\theta)\right) \rho^2 d\rho$
y_p	$\frac{\partial I_2(\theta)}{\partial y_p} = \frac{ka}{z_d} \frac{y_d - y_p}{r(\theta)} \int_0^1 J_1\left(\frac{kar(\theta)\rho}{z_d}\right) \sin\left(W(\rho;\theta)\right) \rho^2 d\rho$
	$\frac{\partial C(\theta)}{\partial x_p} = 0$
	$\frac{\partial I_1(\theta)}{\partial z_p} = -kn_s \int_0^1 \sqrt{1 - \left(\frac{\mathrm{NA}\rho}{n_s}\right)^2} J_0\left(\frac{kar(\theta)\rho}{z_d}\right) \sin\left(W(\rho;\theta)\right) \rho d\rho$
z_p	$\frac{\partial I_2(\theta)}{\partial z_p} = kn_s \int_0^1 \sqrt{1 - \left(\frac{\mathrm{NA}\rho}{n_s}\right)^2} J_0\left(\frac{kar(\theta)\rho}{z_d}\right) \cos\left(W(\rho;\theta)\right) \rho d\rho$
_	$\frac{\partial C(\theta)}{\partial z_p} = 0$
	$\frac{\partial I_1(\theta)}{\partial z_d} = \frac{kar(\theta)}{z_d^2} \int_0^1 J_1\left(\frac{kar(\theta)\rho}{z_d}\right) \cos\left(W(\rho;\theta)\right) \rho^2 d\rho + \frac{ka^2}{2z_d^2} \int_0^1 J_0\left(\frac{kar(\theta)\rho}{z_d}\right) \sin\left(W(\rho;\theta)\right) \rho^3 d\rho$
z_d	$\frac{\partial I_2(\theta)}{\partial z_d} = \frac{kar(\theta)}{z_d^2} \int_0^1 J_1\left(\frac{kar(\theta)\rho}{z_d}\right) \sin\left(W(\rho;\theta)\right) \rho^2 d\rho - \frac{ka^2}{2z_d^2} \int_0^1 J_0\left(\frac{kar(\theta)\rho}{z_d}\right) \cos\left(W(\rho;\theta)\right) \rho^3 d\rho$
	$rac{\partial C(heta)}{\partial z_d} = -rac{4C(heta)}{z_d}$
	$\frac{\partial I_1(\theta)}{\partial A_0} = 0$
A_0	$\frac{\partial I_2(\theta)}{\partial A_0} = 0$
	$\frac{\partial C(\theta)}{\partial A_0} = \frac{2C(\theta)}{A_0}$

_

REFERENCES

- A. V. Abraham, S. Ram, J. Chao, E. S. Ward, and R .J. Ober. Quantitative study of single molecule location estimation techniques. *Opt. Express*, 17(26), December 2009.
- M. Abramowitz and I. A. Stegun, editors. *Handbook of Mathematical Functions*. National Bureau of Standards, 1972.
- F. Aguet. *Super-Resolution Fluorescence Microscopy Based on Physical Models*. PhD thesis, Swiss Federal Institute of Technology Lausanne, EPFL, 2009.
- F. Aguet, D. Van De Ville, and M. Unser. A maximum-likelihood formalism for sub-resolution axial localization of fluorescent nanoparticles. *Optics Express*, 13(26):10503–10522, December 2005.
- E. Betzig, G. H. Patterson, R. Sougrat, O. W. Lindwasser, S. Olenych, J. S Bonifacino, M. W. Davidson, J. Lippincott-Schwartz, and H. F. Hess. Imaging intracellular fluorescent proteins at nanometer resolution. *Science*, 313(5793):1642–1645, September 2006.
- N. Bobroff. Position measurement with a resolution and noise limited instrument. *Rev. Sci. Instrum.*, 57:1152–1157, 1986.
- M. de Moraes Marim, B. Zhang, J. C. Olivo-Marin, and C. Zimmer. Improving single particle localization with an empirically calibrated gaussian kernel. In *Proc. IEEE International Symposium on Biomedical Imaging: From Nano to Macro ISBI*, pages 1003–1006, Paris, France, May 2008.
- S. Geissbühler, C. Dellagiacoma, and T. Lasser. Comparison between SOFI and STORM. *Biomedical Optics Express*, 2(3):408–420, 2011.
- S. Gibson and F. Lanni. Experimental test of an analytical model of aberration in an oilimmersion objective lens used in three-dimensional light microscopy. J. Opt. Soc. Am. A, 9 (1):154–166, January 1992. originally published in J. Opt. Soc. Am. A 8, 1601-1613 (1991).
- S. F. Gibson. *Modeling the 3-D imaging properties of the fluorescence light microscope*. PhD thesis, Carnegie Mellon University, 1990.
- P. N. Hedde, J. Fuchs, F. Oswald, J. Wiedenmann, and G. U. Nienhaus. Online image analysis software for photoactivation localization microscopy. *Nature Methods*, 6(10):689–690, October 2009.
- R. Henriques, M. Lelek, E. F. Fornasiero, F. Valtorta, C. Zimmer, and M. M. Mhlanga.

QuickPALM: 3D real-time photoactivation nanoscopy image processing in ImageJ. *Nature Methods*, 7(5):339–340, May 2010.

- S. T. Hess, T. P. K. Girirajan, and M. D. Mason. Ultra-high resolution imaging by fluorescence photoactivation localization microscopy. *Biophysical Journal*, 91:4258–4272, December 2006.
- B. Huang, W. Wang, M. Bates, and X. Zhuang. Three-dimensional super-resolution imaging by stochastic optical reconstruction microscopy. *Science*, 319(10):810–813, February 2008.
- M. Juette, T. Gould, M. Lessard, M. Mlodzianoski, B. Nagpure, B. Bennet, S. Hess, and J. Bewersdorf. Three-dimensional sub-100 nm resolution fluorescence microscopy of thick samples. *Nature Methods*, 5(6):527–529, June 2008.
- Z. Kam, B. Hanser, M. G. L. Gustafsson, D. A. Agard, and J.W. Sedat. Computational adaptive optics for live three-dimensional biological imaging. *Proc. Natl. Acad. Sci. U.S.A.*, 98(7): 3790–3795, 2001.
- H. Kirshner, T. Pengo, N. Olivier, D. Sage, S. Manley, and M. Unser. A PSF-based approach to biplane calibration in 3D super-resolution microscopy. In *Proceedings of the Ninth IEEE International Symposium on Biomedical Imaging: From Nano to Macro (ISBI'12)*, pages 1232– 1235, Barcelona, Spain, May 2-5, 2012.
- S. Manley, J. M Gillette, G. H. Patterson, H. Shroff, H. F. Hess, E. Betzig, and J. Lippincott-Schwartz. High-density mapping of single-molecule trajectories with photoactivated localization microscopy. *Nature Methods*, 5(2):155–157, 2008.
- J. Markham and J. A. Conchello. Fast maximum-likelihood image-restoration algorithms for three-dimensional fluorescence microscopy. J. Opt. Soc. Am. A, 18(5):1062–1071, 2001.
- I. Märki, N. L. Bocchio, S. Geissbüehler, F. Aguet, A. Bilenca, and T. Lasser. Three-dimensional nano-localization of single fluorescent emitters. *Optics Express*, 18(19):20263–20272, 2010.
- R. J. Ober, S. Ram, and E. S. Ward. Localization accuracy in single-molecule microscopy. *Bio-physical Journal*, 86(2):1185–1200, February 2004.
- Sri Rama Prasanna Pavani and Rafael Piestun. High-efficiency rotating point spread functions. *Opt. Express*, 16(5):3484–3489, Mar 2008.
- P. Prabhat, S. Ram, E.S. Ward, and R.J. Ober. Simultaneous imaging of different focal planes in fluorescence microscopy for the study of cellular dynamics in three dimensions. *IEEE Trans. Nanobioscience*, 3(4):237–242, December 2004.
- C. Preza and J. A. Conchello. Depth-variant maximum-likelihood restoration for three-

dimensional fluorescence microscopy. J. Opt. Soc. Am. A, 21(9):1593-1601, September 2004.

- S. Ram, P. Prabhat, J. Chao, E.S. Ward, and R.J. Ober. High accuracy 3D quantum dot tracking with multifocal plane microscopy for the study of fast intracellular dynamics in live cells. *Biophysical Journal*, 95:6025–6043, December 2008.
- Sripad Ram, E. Sally Ward, and Raimund J. Ober. How accurately can a single molecule be localized in three dimensions using a fluorescence microscope? In Proc. SPIE 5699, Imaging, Manipulation, and Analysis of Biomolecules and Cells: Fundamentals and Applications III, pages 426–435, April 2005. doi: 10.1117/12.587878. URL + http://dx.doi.org/10.1117/12.587878.
- M. J. Rust, M. Bates, and X. Zhuang. Sub-diffraction-limit imaging by stochastic optical reconstruction microscopy (STORM). *Nature Methods*, 3(10):793–796, October 2006.
- J. M. Schmitt and G. Kumar. Turbulent nature of refractive-index variations in biological tissue. *Optics Letters*, 21(16):1310–1312, 1996.
- S. Wolter, M. Schüttpelz, M. Tscherepanow, S. Van De Linde, M. Heilemann, and M. Sauer. Real-time computation of subdiffraction-resolution fluorescence images. *J. Microscopy*, 237 (1):12–22, January 2010.
- B. Zhang, J. Zerubia, and J. C. Olivo-Marin. Gaussian approximations of fluorescence microscope point-spread function models. *Appl- Opt-*, 46(10):1819–1829, April 2007.